

# Melatonin as a Neuroprotective Agent in the Rodent Models of Parkinson's Disease: Is it All Set to Irrefutable Clinical Translation?

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**Abstract** Parkinson's disease (PD), a neurodegenerative disorder, is characterized by the selective degeneration of the nigrostriatal dopaminergic neurons, continuing or permanent deficiency of dopamine, accretion of an abnormal form of alpha synuclein in the adjacent neurons, and dysregulation of ubiquitin proteasomal system, mitochondrial metabolism, permeability and integrity, and cellular apoptosis resulting in rigidity, bradykinesia, resting tremor, and postural instability. Melatonin, an indoleamine produced almost in all the organisms, has anti-inflammatory, anti-apoptotic, and anti-oxidant nature. Experimental studies employing 1-methyl 4-phenyl 1, 2, 3, 6-tetrahydropyridine (MPTP), 6-hydroxydopamine (6-OHDA), methamphetamine, rotenone, and maneb and paraquat models have shown an enormous potential of melatonin in amelioration of the symptomatic features of PD. Although a few reviews published previously have described the multifaceted efficacy of melatonin against MPTP and 6-OHDA rodent models, due to development and validation of the newer models as well as the extensive studies on the usage of melatonin in entrenched PD models, it is worthwhile to bring up to date note on the usage of melatonin as a neuroprotective agent in PD. This article presents an update on the

usage and applications of melatonin in PD models along with incongruous observations. The impending implications in the clinics, success, limitations, and future prospective have also been discussed in this article.

**Keywords** Parkinson's disease · Melatonin · Rodent models · Neuroprotection

## Introduction

Parkinson's disease (PD) is the second most prevalent neurodegenerative disease after Alzheimer's disease. It is characterized by the selective and progressive degeneration of the dopaminergic neurons of the nigrostriatal pathway leading to reduced level of dopamine in the striatum resulting in the onset of various clinical symptoms [1, 2]. While PD is contributed mainly by increasing age, environmental exposure to pesticides and heavy metals on the genetically susceptible individuals largely at the later phase of life is one of the most widely accepted suppositions of its etiology [2]. Rural living, well-water drinking, and farming have been found to be associated with the increased incidences of PD [1]. Environmental chemicals [manganese ethylene bithiocarbamate (maneb), *N,N'*-dimethyl-4,4'-bipyridinium dichloride (paraquat), cypermethrin, rotenone, zinc, lead, iron, manganese, etc.] that preferentially and selectively degenerate the dopaminergic neurons are used to reproduce some of the features of the sporadic PD in the experimental animals. In addition to the environmental chemicals, many drugs, such as reserpine, methamphetamine, 6-hydroxydopamine (6-OHDA), and a contaminant of the synthetic heroin, 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine (MPTP), when administered systemically in the experimental animals,

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produce many symptomatic, neurochemical, and anatomical features mimicking sporadic PD [1–3]. Genome and proteome approaches have identified several genes and proteins, which have offered clues to the genetic origin of PD in some individuals and also paved the way to develop the genetic models to understand its pathogenesis and to assess the efficacy of restorative agents [2].

Melatonin (*N*-acetyl-5-methoxytryptamine), an indoleamine, is a highly conserved anti-oxidant molecule secreted from the pineal gland, gastrointestinal tract, ovaries, testes, bone marrow, and eye lenses [4]. Melatonin and its metabolites, in general, offer anti-inflammatory, anti-apoptotic, anti-oxidative, and free-radical scavenging properties and protect against mitochondrial dysfunction [4–8], thereby regulating multiple biological processes in the body of an organism (Fig. 1). It is known to control the transcription, translation, and catalytic activities of the preventive antioxidants, including glutathione peroxidase, superoxide dismutase (SOD), and catalase under physiological and stress conditions [9]. Melatonin reduces the expression of adhesion molecules and pro-inflammatory cytokines, and regulates the expression of xenobiotic metabolizing enzymes and serum inflammatory indices [4, 8]. Furthermore, melatonin increases the activity of the mitochondrial complex I and complex IV, preserves homeostasis, improves respiration, enhances glutathione level, increases ATP synthesis, and decreases the harmful reduction in the mitochondrial membrane potential, which triggers mitochondrial transition pore opening and the apoptotic cascade [10, 11].

PD is the consequence of divergent contributors, including age and genetic and environmental factors, converging at a mysterious point leading to oxidative stress, inflammation, and selective neuronal death. Despite strong debate and

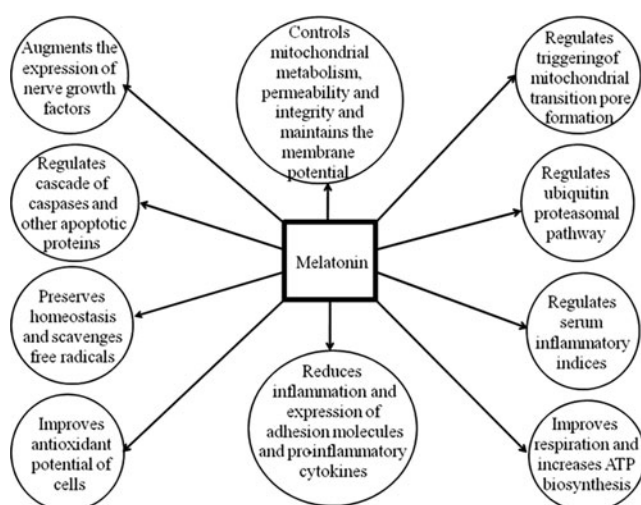
conflicting evidences over various issues on PD, it is widely accepted that the disease is an outcome of an increased oxidative stress leading to irreversible dopaminergic neuronal death and is shared mainly by the impaired mitochondrial integrity and function, energy metabolism, and inefficient ubiquitin proteasomal system [2].

Melatonin offers neuroprotection in a number of neurodegenerative diseases, including PD [7, 8]. Since melatonin is a powerful free-radical scavenger, naturally occurring anti-oxidant defense stimulator, potent anti-apoptotic agent, and a modulator of xenobiotic metabolizing enzymes, therefore, it may be considered to ameliorate the symptomatic features of PD [5, 6, 8, 9]. Melatonin is an ideal neuroprotective agent as it can easily cross the blood–brain barrier and enter the subcellular compartments, lacks toxicity as compared with many other neuroprotective agents, and possesses effective combating efficacy against free-radical-induced neuronal injury [12]. The presence of a considerable amount of melatonin in the mitochondria and growing evidences for mitochondrial dysfunction and oxidative stress in the dopaminergic neurons raise the likelihood of functional implication of melatonin in the mitochondrial and non-mitochondrial functions associated with PD [8, 13]. This is further supported by the fact that melatonin effortlessly enters the brain and cerebrospinal fluid owing to its smaller size and amphiphilic nature [12]. Melatonin is widely accepted as an alternative approach to ameliorate the symptomatic features of PD in experimental animals [8, 14–17].

Because of its ability to inhibit the ubiquitination-dependent proteasomal activity and to regulate numerous neuronal functions by receptor-dependent and receptor-independent mechanisms, melatonin is widely tested for its therapeutic efficacy against established rodent models of PD [7, 8, 16, 18, 19]. However, its wide relevance in clinics for the same purpose is not yet clearly established, and the knowledge generated from epidemiological studies is limited and inconclusive [20, 21]. It is worthwhile to present an update of the information as melatonin is found to be effective even against the recently developed and validated pesticides-induced animal models [1, 8, 22]. This article describes an update on the various rodent models and the interpretation of the usage of melatonin therein, followed by the possibility of its real applications in humans, the challenges associated with its applications, and the possible approaches to overcome these limitations.

## Melatonin and 6-OHDA

6-OHDA is possibly the first chemical reported to degenerate the cell bodies of the dopaminergic neurons in the substantia nigra and fibers in the striatum and also induces microglial activation [23, 24]. It is known to produce several



**Fig. 1** Major biochemical, molecular, and histological events involved in PD, which could be significantly modulated by melatonin



works independent to melatonin, 6-OHDA-induced mitochondrial complex I deficiency, which increases reactive oxygen species production, reduces ATP synthesis, and leads to energy failure, may be taken care of by melatonin [31] (Fig. 2).

Most of the studies have shown that the anti-oxidant, anti-apoptotic, and anti-inflammatory efficacy of melatonin play critical roles in ameliorating symptomatic features of 6-OHDA-induced PD. However, melatonin exacerbates 6-OHDA-mediated motor impairments and cannot work as a universal remedy [34]. Despite rare conflicting reports suggesting the negative effects of melatonin, most of the studies decisively favor that melatonin offers neuroprotection against 6-OHDA [35].

### Melatonin and MPTP

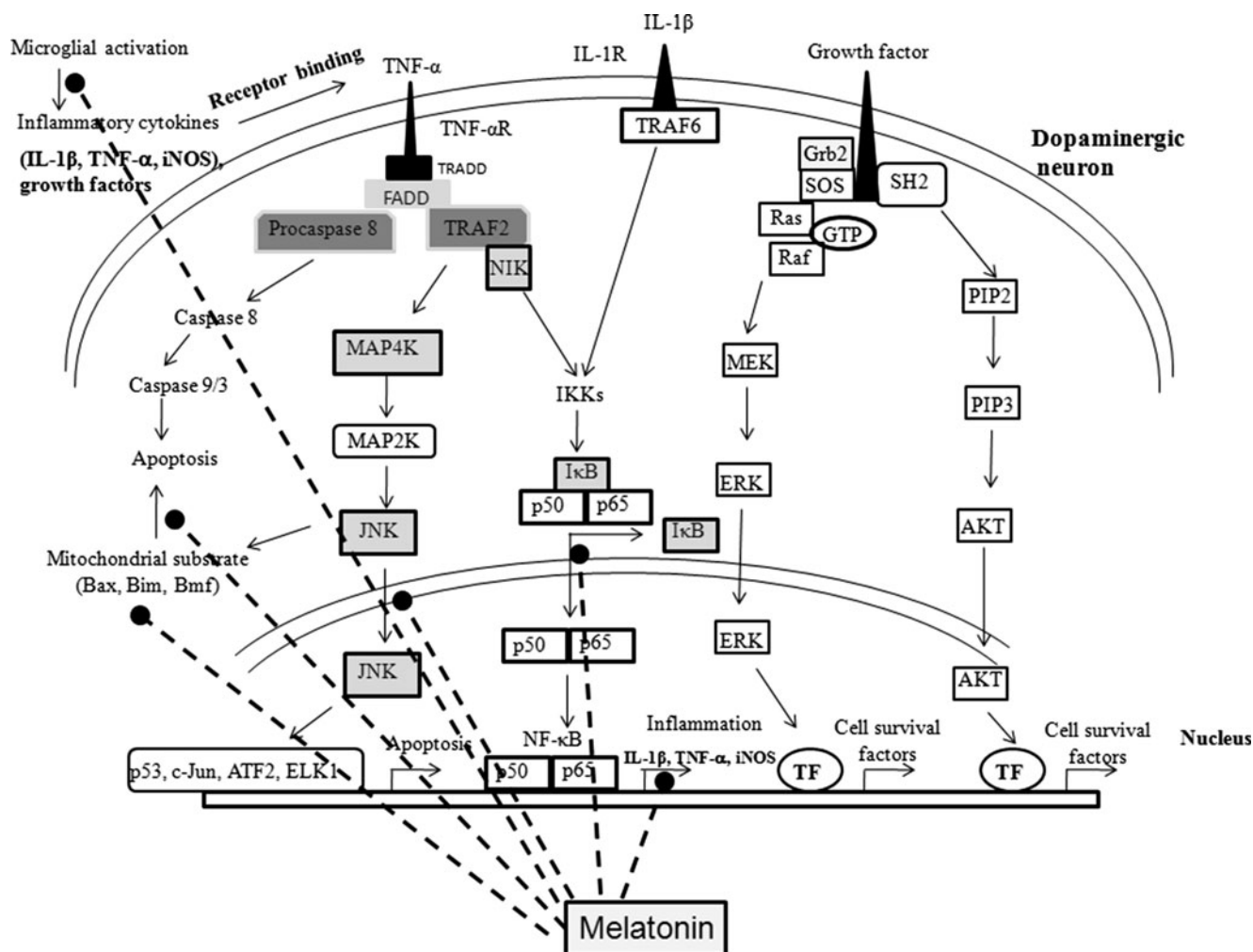
MPTP has been widely used to develop PD symptoms in the experimental animals to understand the pathophysiology and to assess the efficacy of anti-PD drugs. As MPTP is a lipophilic molecule, therefore, it readily crosses the blood–brain barrier [7]. The development of this model system is prompted by the PD phenotype seen during 1980s in the synthetic heroin addicts in North California [36]. It was Langston and his co-workers who identified MPTP, the real culprit, present in the synthetic heroin [37]. MPTP effectively elicits most of the critical features of PD in humans and primates, and a few of them can also be seen in the experimental rodents [7, 38]. Although MPTP does not induce the formation of non-fibrillar Lewy body in the nigrostriatal tissues and many phenotypic symptoms in the rodents, such as resting tremor, it is known to degenerate the dopaminergic neurons by inducing oxidative stress. MPTP induces the free-radical generation owing to its ability to inhibit mitochondrial complex I. MPTP alters nicotinamide adenine dinucleotide phosphate (NADPH) oxidase system in addition to the activation of microglial cells and neuroinflammation [39]. MPTP is known to induce apoptosis via the activation of caspases and augmentation of DNA fragmentation [26, 40] (Fig. 2).

Melatonin, on the other hand, ameliorates the progressive impairment of the mitochondrial function, prophylactically reduces the oxidative damage and lipid peroxidation in MPTP-induced PD [41–43], and protects TH-positive nerve terminals [44, 45]. Since melatonin easily enters the brain, therefore, it inhibits the damage elicited by the chronic administration of MPTP, as measured by the nigral cell count, TH protein level, and other ultra-structural features related to PD pathology [46]. Melatonin is well known to reduce the MPTP-induced increase in the striatal 6-OHDA and protect against dopamine depletion [14]. MPTP-mediated neuroinflammation is a complex event initiated

by the phosphorylation of p38 mitogen-activated protein kinase (p38 MAPK) and translocation of nuclear factor kappa-B (NF- $\kappa$ B) (Fig. 3). Melatonin resists MPTP-induced degeneration of the dopaminergic neurons, nitrosative and oxidative stress, levels of intracellular calcium ion and phosphorylated p38 MAPK, translocation of NF- $\kappa$ B, and release of pro-inflammatory cytokines [47, 48] (Fig. 3). According to a study, the neuroprotective effect of melatonin against MPTP-induced dopaminergic neurotoxicity partially owes to its hydroxyl radical scavenging property [49]. MPTP down-regulates the expression of growth factors; however, melatonin increases the expression of growth factors, such as fibroblast growth factor 9 by up-regulating heme oxygenase-1 and gamma-glutamylcysteine synthetase expressions, thereby resisting the MPTP-mediated toxicity [50]. Melatonin ameliorates MPTP-induced inhibition of the mitochondrial electron transport chain and increase in oxidative damage as well as dysfunction of the glutathione system and blocks caspase-3 activation and cellular apoptosis [51, 52]. Melatonin potentially attenuates MPTP-induced nigrostriatal dopaminergic injury by impeding the increase of oxidized/reduced glutathione ratio owing to its anti-oxidant property [52, 53]. Melatonin counteracts MPTP-induced apoptosis as well as MPTP-dependent DNA fragmentation in vivo [54]. Melatonin also encounters MPTP-induced c-Jun-N-terminal kinase and caspase-dependent signaling leading to the dopaminergic neurodegeneration [55].

Melatonin is reported to produce complete protection against MPTP-induced dopaminergic neurodegeneration even in in vitro system [56]. The major factors responsible for melatonin-mediated neuroprotection against MPTP-induced neurodegeneration are its anti-oxidative, anti-apoptotic, and anti-inflammatory properties and its ability to regulate many associated sequential events. According to a report, melatonin is not a universal remedy and may pose considerable problems in neurological diseases characterized by dopamine depletion [34]; however, its huge potential as a neuroprotective agent and experimental evidences have strongly contradicted the hypothesis. Additionally, some studies claim that melatonin does not inhibit monoamine oxidase-B and therefore could be unable to protect against the MPTP-induced chronic reduction in the striatal dopamine content [57]. Although contradictions have been reported in a few studies, the reasons for such contradictions are difficult to interpret. However, variations may arise because of the time points considered for the isolation and fractionation of the nigrostriatal tissues and mitochondrial preparations under various experimental paradigms. Variations in the results across studies could be maximally minimized when the time of the isolation of tissues and mitochondrial fraction may be taken into account [58].





**Fig. 3** The figure shows toxins-mediated microglial activation, which subsequently release the inflammatory cytokines. These cytokines further bind to respective receptors at the cell surface of the dopaminergic neurons leading to enhanced production of the pro-apoptotic and inflammatory proteins leading to neuronal cell death (solid arrow). The complete signal transduction pathway is mentioned in the image along with the involvement of phosphorylation of various signaling molecules. The effects of melatonin are shown via the dotted lines circled at the end against various events responsible for neurodegeneration (*TNF-α* tumor necrosis factor-α, *TNF-αR* tumor necrosis factor-α receptor, *IL-1β* interleukin-1β, *IL-1R* interleukin-1β receptor, *iNOS* inducible nitric oxide synthase, *SH2* Src homology 2 domain, *FADD* fas-activated death domain, *TRAF 2* tumor necrosis factor-α receptor associated factor 2, *TRAF 6* tumor necrosis factor-α receptor associated factor 6, *MAP2K*

mitogen-activated protein 2 kinase, *MAP4K* mitogen-activated protein 4 kinase, *JNK* c-Jun N-terminal kinase, *IκB* inhibitor of kappa B, *NF-κB* nuclear factor kappa B, *TF* transcription factor, *Bax* B-cell lymphoma 2 associated protein X, *Bim* B-cell lymphoma 2 interacting mediator of cell death, *Bmf* B-cell lymphoma 2 modifying factor, *IKKs* inhibitor of κB kinase, *NIK* nuclear factor-κB inhibitory kinase, *ERK* extracellular signal-related kinase, *Grb2* growth factor receptor-bound protein 2, *SOS* son of sevenless (ras-specific nucleotide-exchange factor), *MEK* MAP kinase kinase, *ATF2* activating transcription factor 2, *ELK 1* E twenty-six like transcription factor 1, *Apaf 1* apoptotic protease activating factor 1, *AKT* Akt transforming serine/threonine protein kinase, *PIP2* phosphatidylinositol diphosphate, *PIP3* phosphatidylinositol triphosphate, *TRADD* tumor necrosis factor receptor type 1-associated death domain)

### Melatonin and Paraquat Either Alone or in Combination with Maneb

Epidemiological evidences have prompted investigators to develop pesticides-induced animal models to understand the molecular and biochemical events leading to PD and to develop therapies to encounter the disease [22, 59, 60]. Paraquat, an herbicide, possesses structural similarity with 1-methyl 4-phenylpyridinium ion (MPP<sup>+</sup>), the primary metabolite of MPTP, and induces the nigrostriatal

dopaminergic neurodegeneration almost in the same fashion as MPTP. Unlike MPTP, paraquat is a polar molecule and crosses the blood–brain barrier through neutral amino acid transporter. Maneb, a fungicide, induces the nigrostriatal dopaminergic neurodegeneration along with paraquat much more potentially than paraquat alone. Paraquat inhibits the mitochondrial complex I; however, mane b inhibits the complex III (Fig. 2). Both of them when administered together lead to free-radical generation, and inhibit proteasomal function and energy metabolism more potentially than

that of alone. These pesticides generate superoxide, hydroxyl, and fatty acyl radicals leading to DNA fragmentation and apoptosis [60–65]. Paraquat also induces microglial activation and inhibits NADPH oxidase activity as observed with MPTP [24, 36].

Melatonin prevents the dopaminergic neurodegeneration by inhibiting free-radical generation, neuroinflammation, and apoptosis induced by paraquat and maneb possibly in the same fashion as in the case of MPTP. Since maneb and paraquat combined model is a relatively newer animal model of PD, the studies on the effect of melatonin against this model system are limited [8]. Melatonin and its metabolite *N* (1)-acetyl-5-methoxykynuramine inhibit the expression and activity of the mitochondrial inducible nitric oxide synthase and prevent mitochondrial failure [17] in many PD models (Fig. 3). Likewise, melatonin reduces the inducible nitric oxide synthase, nitrite content, oxidative stress, and apoptosis in maneb- and paraquat-induced PD phenotype in mouse [8]. Melatonin combats paraquat-induced oxidative stress, regulates the recycling of reduced nicotinamide adenine dinucleotide (NADH), and defends against paraquat-induced NADH depletion [66, 67]. Melatonin checks paraquat-mediated DNA damage and genotoxicity by foraging the hydroxyl and other free radicals [68, 69]. Similarly, melatonin reduces paraquat-induced mortality in rats and increases the lethal dose 50 [70]. Melatonin is found to inhibit the maneb-induced alpha-synuclein aggregation and mitochondrial dysfunctions and neurodegeneration at a night-time physiological blood concentration [71]. Despite little knowledge about the applications of melatonin against maneb- and paraquat-induced toxicity, it can be inferred from the studies conducted so far that melatonin could be equally effective in this model system as in 6-OHDA and MPTP.

### Melatonin and Rotenone

Rotenone, a lipophilic compound, readily crosses the blood–brain barrier, inhibits the mitochondrial electron complex I, causes free-radical generation, and reduces ATP biosynthesis (Fig. 2). Rotenone leads to the dopaminergic neurodegeneration and reduces the number of TH-positive neurons of the nigrostriatal pathway. Although degeneration caused by rotenone is non-specific in nature, it is the only neurotoxin which exhibits well-defined Lewy body formation and aggregation of alpha-synuclein along with depletion of glutathione, disruption of axonal transport, and onset of several critical histological, biochemical, and pathological hallmarks of sporadic PD [23, 27, 72]. Since rotenone does not rely on the dopamine transporter uptake to exert neurotoxicity, therefore, it could be considered as an ideal model to assess neuroprotection [73].

Keeping in view of its close resemblance with the sporadic PD, several studies have been performed to assess the neuroprotective potential of melatonin against this model system. Melatonin prevents the nigrostriatal neurodegeneration and alpha-synuclein aggregation [72] induced by rotenone. Melatonin scavenges rotenone-induced hydroxyl radicals and restores the decreased glutathione level and changes in the catalytic activities of superoxide and catalase in the substantia nigra [74]. Even in chronic rotenone-based *Drosophila* model system, melatonin alleviates both locomotor impairment and the dopaminergic neuronal loss [75]. Although melatonin does not directly alter free calcium ion concentration or rotenone-induced inhibition of mitochondrial complex I, it is known to inhibit calcium ion and rotenone combined-induced oxidative stress in isolated rat brain mitochondria [76] (Fig. 2). Contrary to the above findings, melatonin is reported to potentiate the striatal catecholamine depletion and terminal loss and degeneration of the cell bodies of the nigral dopaminergic neurons, thereby exacerbating many features of rotenone-induced PD in rats [73]. Additionally, melatonin alone elicits an alteration in the striatal catecholamine level showing that melatonin does not offer neuroprotection against rotenone-induced PD in rats [73]. Overall, melatonin offers neuroprotection against rotenone model by inhibiting the free-radical generation, resisting TH-positive neuronal loss, and restoration of the dopamine level in the striatum.

### Melatonin and Methamphetamine/Amphetamine

Methamphetamine-induced striatal dopamine depletion is considered as one of the important models to mimic some of the PD pathology, and chronic or intermittent treatment induces temporary or permanent disturbance in the dopaminergic system [77, 78]. Moreover, methamphetamine increases protein-1 and cyclic adenosine monophosphate response element binding protein expressions by activating the respective transcription factors and also directly acts on the mesencephalic cell nuclei [79]. Amphetamine leads to an increase in the level of alpha-synuclein and decrease in the phosphorylated TH and mitochondrial complex I proteins [15].

The protective effects of melatonin on methamphetamine-induced changes in the nigrostriatal system have been widely studied [78, 80, 81]. Methamphetamine-induced increase in free-radical formation owing to incomplete oxidative phosphorylation and mitochondrial damage leading to a failure of cellular energy metabolism followed by a secondary excitotoxicity could be regulated by melatonin [78] (Fig. 2). It is found that methamphetamine-induced biosynthesis of the reactive nitrogen species, peroxynitrite, which leads to the striatal dopamine depletion, is significantly reduced by melatonin

[80]. Amphetamine-induced changes in alpha-synuclein, phosphorylated TH, and mitochondrial complex I proteins are significantly reversed by melatonin [15]. Melatonin is reported to rescue methamphetamine-induced depletion of the striatal dopamine and dopamine transporter binding sites [82]. Melatonin effectively inhibits JNK 1 activation and reduces apoptotic signaling cascade and autophagic phenotype in methamphetamine-induced autophagy in a dopaminergic neuronal cell line [81] and therefore is useful in treating PD as JNK inhibition can be used to treat PD [83] (Fig. 3). Melatonin counteracts amphetamine-induced loss of cell viability and oxidative stress, and causes a decrease in alpha-synuclein expression and an increase in ATP synthesis in a human dopaminergic neuroblastoma cell line [84]. Melatonin alleviates methamphetamine-induced neurotoxicity, oxidative and nitrosative stress, and also suppresses the expressions of interleukin1-beta, interleukin-6, and tumor necrosis factor-alpha [85]. Additionally, melatonin is reported to protect against methamphetamine-mediated calpain-dependent cell death and increase cell viability [86]. Similarly, melatonin reduces the levels of Bax, Bcl-2, and cleaved caspase 3 proteins and thereby protects against cell death in methamphetamine-treated dopaminergic neuronal cells [87] (Fig. 2). Melatonin activates the mammalian target of rapamycin (mTOR) signaling pathway, phosphorylates mTOR and its downstream target, the eukaryotic initiation factor 4E-binding protein, and inhibits methamphetamine-mediated autophagy in vitro [88]. Melatonin abolishes methamphetamine-induced degeneration of nerve terminals in the neonatal rat brain and partially restores the expressions of TH, synaptophysin, and growth-associated protein-43 [89]. Melatonin protects the dopaminergic and serotonergic neurons against methamphetamine-generated oxidative stress [90]. On the contrary, melatonin is reported to aggravate the methamphetamine-induced deficits in the serotonergic and dopaminergic systems, such as the suppression of the activities of TH and tryptophan hydroxylase and reduction in the contents of dopamine and 5-hydroxytryptamine [91]. Melatonin also reduces the amphetamine-mediated toxic insults in the dopaminergic neurons by preserving the levels of VMAT-2 and phosphorylated TH [92]. Melatonin suppresses the amphetamine-induced overexpression of inducible nitric oxide synthase mRNA and defends against nitrosative stress-induced brain damage [93] (Fig. 3).

### Other Rodent Models and Melatonin

Unfortunately, melatonin is not yet tested against most of the recent PD models, which might have relevance to humans. We are exposed to many chemicals, such as heavy metals and pyrethroid pesticides in our daily life; therefore,

these chemicals are being employed to develop the animal models. Melatonin is not yet checked against metals and cypermethrin-induced nigrostriatal dopaminergic neurodegeneration, as these are relatively newer animal models [94–97].

### Melatonin and Possible Explanations of the Neuroprotective Potential

Free-radical generation, mitochondrial pore formation, and oxidative stress-induced neuronal cell death are critical in PD, and any substance which prevents opening of mitochondrial pores and preserves the mitochondrial function may act as a neuroprotective agent [98]. Melatonin reduces neuroinflammation, oxidative stress, and cell adhesion, and restores mitochondrial function [4,8] and also acts as an important component of the brain's anti-oxidant defense system against catecholamine auto-oxidation and protects against the consequent dopaminergic neurodegeneration [99]. This idea gained momentum from the fact that pineal gland meets the criteria of the neuroendocrine system, and dysfunction of the pineal gland may be associated with the pathophysiology and clinical manifestations of PD [100]. Although initially it was considered that melatonin offers neuroprotection by restoring its level or by improving the restoration of the expression of TH by reducing its oxidation [7], several theories have been documented till date.

### Inhibition of Dopamine Release

An integrated relationship between dopamine and melatonin is essential for normal physiology, and any imbalance between the two may lead to PD [101]. But as the physiological levels of melatonin decrease with age, therefore, its importance in the total antioxidative defense capacity of an organism is being investigated [102]. Melatonin inhibits the release of dopamine in the striatum and limbic system, and decreases the blinking rate in PD patients, suggesting a functional link among blink rate, melatonin secretion, and the striatal dopaminergic functions [103]. PD may show sleep-related symptoms, and a significant improvement in the subjective sleep disturbance by melatonin suggests its relevance in PD [20, 104].

### Antioxidant and Free-Radical Scavenger

As PD imprints long-lasting physiological and pathological permanent effects on the pathogenic proteins, proteasomal machinery, mitochondrial physiology, metabolism, permeability and viability and cellular integrity and apoptosis,

therefore, anti-oxidants may help in the amelioration of the symptomatic features of PD. Melatonin, reported to be a potent free-radical scavenger, forages a variety of reactive oxygen and nitrogen species, including hydroxyl radical, hydrogen peroxide, singlet oxygen, nitric oxide, and peroxynitrite anion. Furthermore, it can improve the cell survival and functions by stimulating anti-oxidant enzymes, increasing the efficiency of the electron transport chain and promoting ATP synthesis [6]. Even the pharmacological levels of melatonin are used to combat oxygen and lipid peroxides-induced toxicities [7]. It is several times more potent than vitamin C and E in protecting tissues from oxidative injury when compared at an equivalent dosage [105]. Melatonin, alone or in combination with deprenyl, defends against the dopaminergic neurodegeneration, as they suppress the hydroxyl radical formation during dopamine autoxidation *in vitro* [106, 107]. Melatonin regulates circadian modulation of Na<sup>+</sup>/K<sup>+</sup>-ATPase and Na<sup>+</sup>/H<sup>+</sup> exchanger owing to its anti-oxidative and membrane fluidity modulating properties [108]. Melatonin-mediated rhythmic modulation of malondialdehyde and intracellular glutathione contents during day and night emphasizes the role of melatonin as an antioxidant and its function against oxidative stress [109].

### Mitochondrial Modulator

As inhibition of mitochondrial complexes I to IV have been implicated in the pathogenesis of PD, therefore, a modulator of any of these, if not all, may be used to protect the dopaminergic neurons from damage. Melatonin preserves mitochondrial homeostasis, enhances mitochondrial glutathione level, and preserves proton potential and ATP synthesis by stimulating complex I and IV activities [11]. Melatonin increases the activity of the complex I and complex IV, and improves mitochondrial respiration, increases ATP synthesis under normal and stressful conditions, and also ameliorates the harmful reduction in the mitochondrial membrane potential that may trigger mitochondrial transition pore opening and the apoptotic cascade [10] (Fig. 2; Table 1). Melatonin protects against the deficits in the mitochondrial complexes leading to free-radical mechanisms, both directly via reactive oxygen species production and indirectly by decreased ATP synthesis and energy failure [31].

### Anti-apoptotic Molecule

Apoptosis is one of the major critical events in the pathophysiology of neurodegenerative diseases, especially PD. Melatonin exhibits the apoptotic gene and protein expression patterns similar to many neuroprotective agents [110].

Melatonin also displays an extremely low index of mortality and even its high concentration does not significantly affect the expression of the mitochondrial Bcl-2 family members, bcl-2 and Bax [98]. Melatonin also inhibits oxidative stress-induced NF- $\kappa$ B activation, one of the main molecular hallmarks of the apoptotic events in PD [111] (Figs. 2 and 3, Tables 1 and 2).

Extracellular signal-regulated kinase (ERK) signaling is involved in the transcription of various genes responsible for cell survival. Melatonin increases the phosphorylation of ERK and activates the mitogen-activated protein kinase (MAPK) pathway in gonadotropin-releasing hormone-secreting neuronal cell line (GT1–7 cells) [112]. Similarly, Akt transforming serine/threonine kinase (Akt) is one of the most important mediators of growth factor-induced cell survival and regulates cell proliferation, apoptosis, and cell-cycle progression [113]. Melatonin is known to induce Akt phosphorylation via melatonin receptor- and phosphatidylinositol-3-phosphate kinase-dependent pathways in primary astrocytes [114], and defends against neuronal death in the hippocampus [115] and cerebral ischemia [116] (Fig. 3).

### Growth Factor Promoter

Melatonin attenuates the compensatory contralateral increase in the striatal glial cell-derived neurotrophic factor (GDNF) expression, supporting a physiological role for melatonin in correcting the expression of growth factors, which is normally defective in PD [117]. Melatonin induces GDNF and melatonin receptor expressions, which supports a functional role for the MT1 receptor, as they are colocalized and their interaction possibly offers neuroprotection against PD [118]. An induction of GDNF mRNA by melatonin also shows how it maintains the nigrostriatal dopaminergic integrity, as GDNF is essential to protect the neurons [119].

### Advantages

Melatonin is an ideal neuroprotective agent in the amelioration of symptomatic features of PD owing to its lesser toxicity, ability to enter the brain, and combating efficacy against free-radical-induced neuronal injury ([12]; Table 2). As PD is mainly an aging-related disease characterized by an increased oxidative stress and reduced or altered expression of anti-oxidant, anti-inflammatory, toxicant metabolizing, apoptotic, and energy metabolism-related genes, and melatonin possesses the potent anti-oxidant, free-radical scavenging, anti-apoptotic, and anti-inflammatory properties, therefore it can effectively encounter the altered expression of several genes and proteins



**Table 1** Summary of a few studies showing the major effects of melatonin against chemicals-induced cellular and animal PD models

Animal model	Salient features	Effects of melatonin	References
6-OHDA	Inhibits the mitochondrial complexes I and IV, causes non-selective degeneration of the dopaminergic neurons and produces Lewy body	Inhibits neuroinflammation and protects the dopaminergic neurons by restoring complex I activity, reduces microglial activation, apomorphine-induced rotations, oxidative stress and attenuates apoptosis	[7, 30–32]
MPTP	Inhibits the mitochondrial complex I and causes selective, however, mainly acute degeneration of dopaminergic neurons and does not form distinct Lewy body	Inhibits neuroinflammation and 6-OHDA production, protects the dopaminergic neurons by reducing TH-positive cell loss and hypolocomotion, restores mitochondrial complex I activity, decreases microglial activation, attenuates mitochondrial DNA damage, prevents dysfunction of glutathione system, blocks caspase-3 activation, protects the nigrostriatum from oxidative stress, and reduces oxidative stress and lipid peroxidation	[14, 41, 44–49, 51–53]
Rotenone	Inhibits complex I of the electron transport chain, non-selectively degenerates the dopaminergic neurons, induces acute neurodegeneration and causes Lewy body formation	Prevents the nigrostriatal neurodegeneration and alpha-synuclein aggregation, reduces glutathione depletion and catalase activity, increases SOD activity, protects the mitochondria from oxidative stress, and increases cell viability	[72, 74, 76]
Maneb and paraquat	Inhibit complex I and III and degenerate the dopaminergic neurons	Protects the dopaminergic neurons and inhibits the expression of pro-apoptotic genes	[8]
Methamphetamine/amphetamine	Degenerates the dopaminergic neurons and reduces dopamine packaging in synaptic vesicles	Attenuates VMAT-2 reduction and inhibits inducible nitric oxide synthase expression, increase in alpha-synuclein, decrease in phosphorylated TH and the mitochondrial complex I proteins	[15, 84, 92, 93]

**Table 2** Tabular representation of the possible therapeutic applications of melatonin and the plausible limitations of therapy based on the knowledge gathered from various experimental models of PD

Animal model	Potential therapeutic applications	Limitations of therapeutic applications	References
6-OHDA	Protects against oxidative stress-induced apoptosis and restores the striatal dopaminergic function and behavioral deficits	Any modulation in the basal level of regular apoptosis must be checked Possibly exacerbate behavioral deficits and impairment of motor function	[28–30, 33, 34]
MPTP	Protects TH immunoreactivity of nerve terminals against reactive oxygen species and oxidative stress-induced apoptosis, and prevents the dopamine depletion in the striatum, PD progression, and the interaction of MPP <sup>+</sup> and other toxins with the mitochondrial complex I	Perturbation in the physiological balance between the body fluid concentrations of melatonin and dopamine owing to exogenous melatonin delivery may exacerbate neurodegeneration Stage-specific response needs elucidation Whether interaction is prohibited in vivo, irrespective of the route of exposure, needs to be sorted out	[14, 41, 44, 46, 47, 51, 101]
Maneb and paraquat	Resists oxidative stress-induced neurodegeneration and inhibits the aggregation of alpha-synuclein and mitochondrial dysfunction	Any modulation in the basal level of regular apoptosis must be checked Extrapolation of the similar result in human subjects needs validation	[8, 71]
Rotenone	Restores antioxidant enzymes in the substantia nigra and prevents the nigrostriatal neurodegeneration	Possibly potentiate the striatal catecholamine depletion and degeneration of the nigral dopaminergic neurons	[72–74]
Methamphetamine/amphetamine	Encounters oxidative and nitrosative stress, subsequent autophagy and apoptosis and restores native conformation and structure of PD-related proteins, such as alpha-synuclein, TH, and mitochondrial complex I	Any modulation in the basal level of regular autophagy and apoptosis must be checked Melatonin may aggravate the disease	[15, 34, 78, 80, 81]

critical in PD pathogenesis [7, 8, 12, 35]. Since melatonin is naturally present in all organisms, could resist multiple anomalies associated with PD unlike most of the anti-PD molecules, and is a potent modulator of xenobiotic metabolizing enzymes, therefore, it has immense potential to be a remedy for PD. Melatonin may be considered as a “wonder” or “many in one” molecule if it succeeds substantially in epidemiological and clinical investigations globally.

The role of melatonin, a potent endogenous antioxidant, has been widely validated not only on the basis of its neuroprotective efficacy assessment but also because of the down-regulation of melatonin receptors in the nigrostriatal region of PD brain [120]. Although melatonin may cause a few side effects, which include nausea, headache, nightmares, and reduced blood flow after high doses and long-term exposures, according to most of the reports, melatonin is totally safe with short-term usage [121, 122]. Despite a report based on a case–control study, which has shown that melatonin improves the subjective quality of sleep but does not restore or improve the motor dysfunction, melatonin is an ideal remedy that can be tested in clinics. The negative result could be probably because of small sample sizes used in the study [21].

Regular melatonin intake by aged individuals may protect from the aging-related depredation, but the results of such a therapy would not be known possibly for decades [123]. The impact of application of melatonin in neurodegenerative disease may get certification only after clinical disease course outcome data become available [124]. Successful scientific evidences through comprehensive human population-based studies will provide real application of melatonin against PD in clinics.

## Limitations

Despite neuroprotective experimental evidences of melatonin, clinical works with melatonin did not observe a remarkable therapeutic efficacy, and a few preclinical and clinical works have also shown that melatonin may even worsen the disease condition [34, 101]. Melatonin has been validated as a neuroprotective agent against most of the well-established animal models of PD (Table 1); studies have also shown that it offers an excellent neuroprotective efficacy against Alzheimer’s disease but not PD [11]. Similarly, multiple effects of melatonin against several complicated disorders may create problems for its proper, complication-free, and widespread usage against PD. For example, a lower rate of cancer mortality or incidence in PD patients speculates the risk or preventative factors common to both diseases, including the lifestyle factors and genetic susceptibility. Lower melatonin concentration predicts a higher cancer and lower PD risk [125]. Application of high doses and long-term exposures of melatonin have been associated with a few minor complications (Table 2), although no major problems have

been associated with its usage so far to the best of our knowledge.

The major causes of limitation of melatonin usage in clinics have been the elusive onset and pathogenesis of the disease and lack of suitable fingerprints for its early diagnosis. PD is mainly a sporadic or idiopathic disorder, and the pathogenesis is often slow and progressive. Since the disease is characterized by the irreversible degeneration of dopaminergic neurons, therefore, the permanent cure of PD is not yet possible with any known therapies. Similarly, none of the animal models developed so far completely or substantially mimics sporadic PD, which indicates that the conclusions drawn from animal experimentations may or may not be completely reproduced in clinical interventions. Melatonin plays a dual role in homeostasis and disease etiology, thereby requiring appropriate assessment of PD pathogenesis in a proper model system employing multiple biochemical, histological, molecular, and phenotypic parameters. Like all other drugs used in PD till date, melatonin does not regenerate the dopaminergic neurons in the nigrostriatal pathway; therefore, the permanent cure using melatonin as a gold standard is also not feasible. Melatonin is an important molecule and possibly has a great future in PD research, but it needs to be extensively tested across multiple populations for efficacy and real effects along with the side effects at the efficacious doses, and only if everything goes alright, it may be considered worthwhile in clinics.

## Future Directions

Melatonin has been recommended for testing against the onset or progression of PD because of its multiple beneficial effects, lack of toxicity, and endogenous nature [7]. However, either its wide application as a therapeutic agent to encounter sporadic PD is not widely conducted or such information is not extensively reported in peer-reviewed, highly rated journals. It is essential to monitor the effects of melatonin in the chronic PD models mimicking sporadic PD more appropriately, employing behavioral, biochemical, histological, ultra-structural, genetic, molecular, genomic, and proteomic tools. Such testing is relevant and may be performed for better doable application of this endogenous molecule against a disease with an elusive pathophysiology and no permanent cure.

Complimentary and combination therapies, such as use of melatonin with dopaminergic therapy, offer more relevant and potent effects on motor and non-motor symptoms; however, clinicians need to ensure that complementary and combinational therapies should be used appropriately without reducing the benefits of each other [126]. Melatonin in combination with stem cell transplantation is found to be more effective as compared with any of them alone [127].

As stem cell therapy is projected to treat PD after successful animal experimentations, its combination with melatonin could be a viable approach to restore normal histopathology and biochemistry in PD patients [127].

Owing to multiple assets of melatonin that include anti-oxidant and free-radical-scavenging properties, anti-inflammatory potential, and its beneficial effects against mitochondrial dysfunction, its metabolites and synthetic analogs need to be tested for highest activity and lowest side effects [128–130]. If such strategies succeed, best melatonin analog could be tested either alone or in combination with other potential therapies to achieve the maximum neuroprotection and minimum toxicity in the sufferers of PD. Endogenous melatonin protects exogenous dopamine precursor from auto-oxidation, and their systemic co-administration increases its bioavailability and long-term application possibly without measurable toxicity. A cocktail therapy of multiple neuroprotective agents that possess lesser toxicities can possibly offer more neuroprotection and rescue the dopaminergic neurons more efficiently than the current treatment strategies with extremely high doses of a single anti-oxidant in the early stages of PD [131].

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## References

- Singh MP, Patel S, Dikshit M, Gupta YK (2006) Contribution of genomics and proteomics in understanding the role of modifying factors in Parkinson's disease. *Indian J Biochem Biophys* 43:69–81
- Srivastava G, Singh K, Tiwari MN, Singh MP (2010) Proteomics in Parkinson's disease: current trends, translational snags and future possibilities. *Expert Rev Proteomics* 7:127–139
- Betarbet R, Sherer TB, Greenamyre JT (2002) Animal models of Parkinson's disease. *Bioessays* 24:308–318
- Esposito E, Cuzzocrea S (2010) Antiinflammatory activity of melatonin in central nervous system. *Curr Neuropharmacol* 8:228–242
- Galano A, Tan DX, Reiter RJ (2011) Melatonin as a natural ally against oxidative stress: a physicochemical examination. *J Pineal Res* 51:1–16
- Leon J, Acuna-Castroviejo D, Sainz RM, Mayo JC, Tan DX, Reiter RJ (2004) Melatonin and mitochondrial function. *Life Sci* 75:765–790
- Mayo JC, Sainz RM, Tan DX, Antolin I, Rodríguez C, Reiter RJ (2005) Melatonin and Parkinson's disease. *Endocrine* 27:169–178
- Singhal NK, Srivastava G, Patel DK, Jain SK, Singh MP (2011) Melatonin or silymarin reduces maneb- and paraquat-induced Parkinson's disease phenotype in the mouse. *J Pineal Res* 50:97–109
- Rodríguez C, Mayo JC, Sainz RM, Antolin I, Herrera F, Martín V, Reiter RJ (2004) Regulation of antioxidant enzymes: a significant role for melatonin. *J Pineal Res* 36:1–9
- Leon J, Acuna-Castroviejo D, Escames G, Tan DX, Reiter RJ (2005) Melatonin mitigates mitochondrial malfunction. *J Pineal Res* 38:1–9
- Srinivasan V, Pandi-Perumal SR, Maestroni GJ, Esquifino AI, Hardeland R, Cardinali DP (2005) Role of melatonin in neurodegenerative diseases. *Neurotox Res* 7:293–318
- Gupta YK, Gupta M, Kohli K (2003) Neuroprotective role of melatonin in oxidative stress vulnerable brain. *Indian J Physiol Pharmacol* 47:373–386
- Escames G, Lopez A, Garcia JA, Garcia L, Acuna-Castroviejo D, Garcia JJ, Lopez LC (2010) The role of mitochondria in brain aging and the effects of melatonin. *Curr Neuropharmacol* 8:182–193
- Borah A, Mohanakumar KP (2009) Melatonin inhibits 6-hydroxydopamine production in the brain to protect against experimental Parkinsonism in rodents. *J Pineal Res* 47:293–300
- Klongpanichapak S, Phansuwan-Pujito P, Ebadi M, Govitrapong P (2008) Melatonin inhibits amphetamine-induced increase in alpha-synuclein and decrease in phosphorylated tyrosine hydroxylase in SK-N-SH cells. *Neurosci Lett* 436:309–313
- Reiter RJ, Tan DX, Jou MJ, Korkmaz A, Manchester LC, Paredes SD (2008) Biogenic amines in the reduction of oxidative stress: melatonin and its metabolites. *Neuro Endocrinol Lett* 29:391–398
- Tapias V, Escames G, López LC, Lopez A, Camacho E, Carrión MD, Entrena A, Gallo MA, Espinosa A, Acuña-Castroviejo D (2009) Melatonin and its brain metabolite N(1)-acetyl-5-methoxykynuramine prevent mitochondrial nitric oxide synthase induction in parkinsonian mice. *J Neurosci Res* 87:3002–3010
- Jan JE, Reiter RJ, Wong PK, Bax MC, Ribary U, Wasdell MB (2011) Melatonin has membrane receptor-independent hypnotic action on neurons: an hypothesis. *J Pineal Res* 50:233–240
- Kwon KJ, Kim JN, Kim MK, Lee J, Ignarro LJ, Kim HJ, Shin CY, Han SH (2011) Melatonin synergistically increases resveratrol-induced heme oxygenase-1 expression through the inhibition of ubiquitin-dependent proteasome pathway: a possible role in neuroprotection. *J Pineal Res* 50:110–123
- Catala MD, Canete-Nicolas C, Iradi A, Tarazona PJ, Tormos JM, Pascual-Leone A (1997) Melatonin levels in Parkinson's disease: drug therapy versus electrical stimulation of the internal globus pallidus. *Exp Gerontol* 32:553–558
- Medeiros CA, Carvalhede de Bruin PF, Lopes LA, Magalhaes MC, de Lourdes SM, de Bruin VM (2007) Effect of exogenous melatonin on sleep and motor dysfunction in Parkinson's disease. A randomized, double blind, placebo-controlled study. *J Neurol* 254:459–464
- Thiruchelvam M, Richfield EK, Goodman BM, Baggs RB, Cory-Slechta DA (2002) Developmental exposure to the pesticides paraquat and maneb and the Parkinson's disease phenotype. *Neurotoxicology* 23:621–633
- Bove J, Prou D, Perier C, Przedborski S (2005) Toxin-induced models of Parkinson's disease. *NeuroRx* 2:484–494
- Croisier E, Moran LB, Dexter DT, Pearce RK, Graeber MB (2005) Microglial inflammation in the parkinsonian substantia nigra: relationship to alpha-synuclein deposition. *J Neuroinflammation* 2:14
- Glinka Y, Gassen M, Youdim MB (1997) Mechanism of 6-hydroxydopamine neurotoxicity. *J Neural Transm Suppl* 50:55–66
- Schober A (2004) Classic toxin-induced animal models of Parkinson's disease: 6-OHDA and MPTP. *Cell Tissue Res* 318:215–224
- Uversky VN (2004) Neurotoxicant-induced animal models of Parkinson's disease: understanding the role of rotenone, maneb and paraquat in neurodegeneration. *Cell Tissue Res* 318:225–241
- Mayo JC, Sainz RM, Antolin I, Rodríguez C (1999) Ultrastructural confirmation of neuronal protection by melatonin against the

- neurotoxin 6-hydroxydopamine cell damage. *Brain Res* 818:221–227
29. Kim YS, Joo WS, Jin BK, Cho YH, Baik HH, Park CW (1998) Melatonin protects 6-OHDA-induced neuronal death of nigrostriatal dopaminergic system. *Neuroreport* 9:2387–2390
  30. Joo WS, Jin BK, Park CW, Maeng SH, Kim YS (1998) Melatonin increases striatal dopaminergic function in 6-OHDA-lesioned rats. *Neuroreport* 9:4123–4126
  31. Dabbeni-Sala F, Di Santo S, Franceschini D, Skaper SD, Giusti P (2001) Melatonin protects against 6-OHDA-induced neurotoxicity in rats: a role for mitochondrial complex I activity. *FASEB J* 15:164–170
  32. Aguiar LM, Vasconcelos SM, Sousa FC, Viana GS (2002) Melatonin reverses neurochemical alterations induced by 6-OHDA in rat striatum. *Life Sci* 70:1041–1051
  33. Mayo JC, Sainz RM, Uria H, Antolin I, Esteban MM, Rodriguez C (1998) Melatonin prevents apoptosis induced by 6-hydroxydopamine in neuronal cells: implications for Parkinson's disease. *J Pineal Res* 24:179–192
  34. Willis GL, Armstrong SM (1999) A therapeutic role for melatonin antagonism in experimental models of Parkinson's disease. *Physiol Behav* 66:785–795
  35. Singh S, Ahmed R, Sagar RK, Krishana B (2006) Neuroprotection of the nigrostriatal dopaminergic neurons by melatonin in hemiparkinsonism rat. *Indian J Med Res* 124:419–426
  36. Miller RL, James-Kracke M, Sun GY, Sun AY (2009) Oxidative and inflammatory pathways in Parkinson's disease. *Neurochem Res* 34:55–65
  37. Langston JW, Ballard P, Tetrud JW, Irwin I (1983) Chronic Parkinsonism in humans due to a product of meperidine-analog synthesis. *Science* 219:979–980
  38. Langston JW, Forno LS, Tetrud J, Reeves AG, Kaplan JA, Karluk D (1999) Evidence of active nerve cell degeneration in the substantia nigra of humans years after 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine exposure. *Ann Neurol* 46:598–605
  39. Gao HM, Liu B, Zhang W, Hong JS (2003) Critical role of microglial NADPH oxidase-derived free radicals in the in vitro MPTP model of Parkinson's disease. *FASEB J* 17:1954–1956
  40. Kotake Y, Ohta S (2003) MPP<sup>+</sup> analogs acting on mitochondria and inducing neuro-degeneration. *Curr Med Chem* 10:2507–2516
  41. Absi E, Ayala A, Machado A, Parrado J (2000) Protective effect of melatonin against the 1-methyl-4-phenylpyridinium-induced inhibition of complex I of the mitochondrial respiratory chain. *J Pineal Res* 29:40–47
  42. Jin BK, Shin DY, Jeong MY, Gwag MR, Baik HW, Yoon KS, Cho YH, Joo WS, Kim YS, Baik HH (1998) Melatonin protects nigral dopaminergic neurons from 1-methyl-4-phenylpyridinium (MPP<sup>+</sup>) neurotoxicity in rats. *Neurosci Lett* 245:61–64
  43. Reiter RJ (1998) Oxidative damage in the central nervous system: protection by melatonin. *Prog Neurobiol* 56:359–384
  44. Acuna-Castroviejo D, Coto-Montes A, Gaia Monti M, Ortiz GG, Reiter RJ (1997) Melatonin is protective against MPTP-induced striatal and hippocampal lesions. *Life Sci* 60:PL23–PL29
  45. Capitelli C, Sereniki A, Lima MM, Reksidler AB, Tufik S, Vital MA (2008) Melatonin attenuates tyrosine hydroxylase loss and hypolocomotion in MPTP-lesioned rats. *Eur J Pharmacol* 594:101–108
  46. Antolín I, Mayo JC, Sainz RM, del Brío ML, Herrera F, Martín V, Rodríguez C (2002) Protective effect of melatonin in a chronic experimental model of Parkinson's disease. *Brain Res* 943:163–173
  47. Ma J, Shaw VE, Mitrofanis J (2009) Does melatonin help save dopaminergic cells in MPTP-treated mice? *Parkinsonism Relat Disord* 15:307–314
  48. Niranjana R, Nath C, Shukla R (2010) The mechanism of action of MPTP-induced neuroinflammation and its modulation by melatonin in rat astrocytoma cells, C6. *Free Radic Res* 44:1304–1316
  49. Li XJ, Gu J, Lu SD, Sun FY (2002) Melatonin attenuates MPTP-induced dopaminergic neuronal injury associated with scavenging hydroxyl radical. *J Pineal Res* 32:47–52
  50. Huang JY, Chuang JI (2010) Fibroblast growth factor 9 upregulates heme oxygenase-1 and gamma-glutamylcysteine synthetase expression to protect neurons from 1-methyl-4-phenylpyridinium toxicity. *Free Radic Biol Med* 49:1099–1108
  51. Chuang JI, Chen TH (2004) Effect of melatonin on temporal changes of reactive oxygen species and glutathione after MPP<sup>+</sup> treatment in human astrocytoma U373MG cells. *J Pineal Res* 36:117–125
  52. Thomas B, Mohanakumar KP (2004) Melatonin protects against oxidative stress caused by 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine in the mouse nigrostriatum. *J Pineal Res* 36:25–32
  53. Chen ST, Chuang JI, Hong MH, Li EI (2002) Melatonin attenuates MPP<sup>+</sup>-induced neurodegeneration and glutathione impairment in the nigrostriatal dopaminergic pathway. *J Pineal Res* 32:262–269
  54. Ortiz GG, Crespo-López ME, Moran-Moguel C, García JJ, Reiter RJ, Acuna-Castroviejo D (2001) Protective role of melatonin against MPTP-induced mouse brain cell DNA fragmentation and apoptosis in vivo. *Neuro Endocrinol Lett* 22:101–108
  55. Chetsawang J, Govitrapong P, Chetsawang B (2007) Melatonin inhibits MPP<sup>+</sup>-induced caspase-mediated death pathway and DNA fragmentation factor-45 cleavage in SK-N-SH cultured cells. *J Pineal Res* 43:115–120
  56. Iacovitti L, Stull ND, Johnston K (1997) Melatonin rescues dopamine neurons from cell death in tissue culture models of oxidative stress. *Brain Res* 768:317–326
  57. Van der Schyf CJ, Castagnoli K, Palmer S, Hazelwood L, Castagnoli N Jr (2000) Melatonin fails to protect against long-term MPTP-induced dopamine depletion in mouse striatum. *Neurotox Res* 1:261–269
  58. Simon N, Papa K, Vidal J, Boulamery A, Bruguerolle B (2003) Circadian rhythms of oxidative phosphorylation: effects of rotenone and melatonin on isolated rat brain mitochondria. *Chronobiol Int* 20:451–461
  59. McCormack AL, Thiruchelvam M, Manning-Bog AB, Thiffault C, Langston JW, Cory-Slechta DA, Di Monte DA (2002) Environmental risk factors and Parkinson's disease: selective degeneration of nigral dopaminergic neurons caused by the herbicide paraquat. *Neurobiol Dis* 10:119–217
  60. Patel S, Singh V, Kumar A, Gupta YK, Singh MP (2006) Status of antioxidant defense system and expression of toxicant responsive genes in striatum of maneb- and paraquat-induced Parkinson's disease phenotype in mouse: mechanism of neurodegeneration. *Brain Res* 1081:9–18
  61. Patel S, Sinha A, Singh MP (2007) Identification of differentially expressed proteins in striatum of maneb- and paraquat-induced Parkinson's disease phenotype in mouse. *Neurotoxicol Teratol* 29:578–585
  62. Patel S, Singh K, Singh S, Singh MP (2008) Gene expression profiles of mouse striatum in control and maneb + paraquat-induced Parkinson's disease phenotype: validation of differentially expressed energy metabolizing transcripts. *Mol Biotechnol* 40:59–68
  63. Peng J, Stevenson FF, Doctrow SR, Andersen JK (2005) Super-oxide dismutase/catalase mimetics are neuroprotective against selective paraquat-mediated dopaminergic neuron death in the substantia nigra: implications for Parkinson disease. *J Biol Chem* 280:29194–29198
  64. Shimizu K, Matsubara K, Ohtaki K, Shiono H (2003) Paraquat leads to dopaminergic neural vulnerability in organotypic midbrain culture. *Neurosci Res* 46:523–532
  65. Thiruchelvam M, Richfield EK, Baggs RB, Tank AW, Cory-Slechta DA (2000) The nigrostriatal dopaminergic system as a



- preferential target of repeated exposures to combined paraquat and maneb: implications for Parkinson's disease. *J Neurosci* 20:9207–9214
66. Bonilla E, Medina-Leendertz S, Villalobos V, Molero L, Bohorquez A (2006) Paraquat-induced oxidative stress in *Drosophila melanogaster*: effects of melatonin, glutathione, serotonin, minocycline, lipoic acid and ascorbic acid. *Neurochem Res* 31:1425–1432
  67. Tan DX, Manchester LC, Sainz RM, Mayo JC, Leon J, Hardeland R, Poeggeler B, Reiter RJ (2005) Interactions between melatonin and nicotinamide nucleotide: NADH preservation in cells and in cell-free systems by melatonin. *J Pineal Res* 39:185–194
  68. Melchiorri D, Ortiz GG, Reiter RJ, Sewerynek E, Daniels WM, Pablos MI, Nisticò G (1998) Melatonin reduces paraquat-induced genotoxicity in mice. *Toxicol Lett* 95:103–108
  69. Yamamoto HA, Mohanan PV (2001) Effects of melatonin on paraquat or ultraviolet light exposure-induced DNA damage. *J Pineal Res* 31:308–313
  70. Melchiorri D, Reiter RJ, Sewerynek E, Hara M, Chen L, Nistico G (1996) Paraquat toxicity and oxidative damage. Reduction by melatonin. *Biochem Pharmacol* 51:1095–1099
  71. Ishido M (2007) Melatonin inhibits maneb-induced aggregation of alpha-synuclein in rat pheochromocytoma cells. *J Pineal Res* 42:125–130
  72. Lin CH, Huang JY, Ching CH, Chuang JI (2008) Melatonin reduces the neuronal loss, downregulation of dopamine transporter, and upregulation of D2 receptor in rotenone-induced parkinsonian rats. *J Pineal Res* 44:205–213
  73. Tapias V, Cannon JR, Greenamyre JT (2010) Melatonin treatment potentiates neurodegeneration in a rat rotenone Parkinson's disease model. *J Neurosci Res* 88:420–427
  74. Saravanan KS, Sindhu KM, Mohanakumar KP (2007) Melatonin protects against rotenone-induced oxidative stress in a hemiparkinsonian rat model. *J Pineal Res* 42:247–253
  75. Coulom H, Birman S (2004) Chronic exposure to rotenone models sporadic Parkinson's disease in *Drosophila melanogaster*. *J Neurosci* 24:10993–10998
  76. Sousa SC, Castilho RF (2005) Protective effect of melatonin on rotenone plus Ca<sup>2+</sup>-induced mitochondrial oxidative stress and PC12 cell death. *Antioxid Redox Signal* 7:1110–1116
  77. Gerlach M, Reiderer P (1996) Animal models of Parkinson's disease: an empirical comparison with the phenomenology of the disease in man. *J Neural Transm* 103:987–1041
  78. Virmani A, Gaetani F, Imam S, Binienda Z, Ali S (2002) The protective role of L-carnitine against neurotoxicity evoked by drug of abuse, methamphetamine, could be related to mitochondrial dysfunction. *Ann N Y Acad Sci* 965:225–232
  79. Asanuma M, Hayashi T, Ordóñez SV, Ogawa N, Cadet JL (2000) Direct interactions of METH with the nucleus. *Mol Brain Res* 80:237–243
  80. Imam SZ, el-Yazal J, Newport GD, Itzhak Y, Cadet JL, Slikker W Jr, Ali SF (2001) Methamphetamine-induced dopaminergic neurotoxicity: role of peroxynitrite and neuroprotective role of antioxidants and peroxynitrite decomposition catalysts. *Ann N Y Acad Sci* 939:366–380
  81. Nopparat C, Porter JE, Ebadi M, Govitrapong P (2010) The mechanism for the neuroprotective effect of melatonin against methamphetamine-induced autophagy. *J Pineal Res* 49:382–389
  82. Itzhak Y, Martin JL, Black MD, Ali SF (1998) Effect of melatonin on methamphetamine- and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced dopaminergic neurotoxicity and methamphetamine-induced behavioral sensitization. *Neuropharmacology* 37:781–791
  83. Wang W, Ma C, Mao Z, Li M (2004) JNK inhibition as a potential strategy in treating Parkinson's disease. *Drug News Perspect* 17:646–654
  84. Klongpanichapak S, Phansuwan-Pujito P, Ebadi M, Govitrapong P (2007) Melatonin protects SK-N-SH neuroblastoma cells from amphetamine-induced neurotoxicity. *J Pineal Res* 43:65–73
  85. Tocharus J, Khonthun C, Chongthammakun S, Govitrapong P (2010) Melatonin attenuates methamphetamine-induced overexpression of pro-inflammatory cytokines in microglial cell lines. *J Pineal Res* 48:347–352
  86. Suwanjang W, Phansuwan-Pujito P, Govitrapong P, Chetsawang B (2010) The protective effect of melatonin on methamphetamine-induced calpain-dependent death pathway in human neuroblastoma SH-SY5Y cultured cells. *J Pineal Res* 48:94–101
  87. Wisessmith W, Phansuwan-Pujito P, Govitrapong P, Chetsawang B (2009) Melatonin reduces induction of Bax, caspase and cell death in methamphetamine-treated human neuroblastoma SH-SY5Y cultured cells. *J Pineal Res* 46:433–440
  88. Kongsuphol P, Mukda S, Nopparat C, Villarroel A, Govitrapong P (2009) Melatonin attenuates methamphetamine-induced deactivation of the mammalian target of rapamycin signaling to induce autophagy in SK-N-SH cells. *J Pineal Res* 46:199–206
  89. Kaewsuk S, Sae-ung K, Phansuwan-Pujito P, Govitrapong P (2009) Melatonin attenuates methamphetamine-induced reduction of tyrosine hydroxylase, synaptophysin and growth-associated protein-43 levels in the neonatal rat brain. *Neurochem Int* 55:397–405
  90. Hirata H, Asanuma M, Cadet JL (1998) Melatonin attenuates methamphetamine-induced toxic effects on dopamine and serotonin terminals in mouse brain. *Synapse* 30:150–155
  91. Gibb JW, Bush L, Hanson GR (1997) Exacerbation of methamphetamine-induced neurochemical deficits by melatonin. *J Pharmacol Exp Ther* 283:630–635
  92. Mukda S, Vimolratana O, Govitrapong P (2011) Melatonin attenuates the amphetamine-induced decrease in vesicular monoamine transporter-2 expression in postnatal rat striatum. *Neurosci Lett* 488:154–157
  93. Tocharus J, Chongthammakun S, Govitrapong P (2008) Melatonin inhibits amphetamine-induced nitric oxide synthase mRNA overexpression in microglial cell lines. *Neurosci Lett* 439:134–137
  94. Kumar A, Ahmad I, Shukla S, Singh BK, Patel DK, Pandey HP, Singh C (2010) Effect of zinc and paraquat co-exposure on neurodegeneration: modulation of oxidative stress and expression of metallothioneins, toxicant responsive and transporter genes in rats. *Free Radic Res* 44:950–965
  95. Singh AK, Tiwari MN, Upadhyay G, Patel DK, Singh D, Prakash O, Singh MP (2012) Long-term exposure to cypermethrin induces the nigrostriatal dopaminergic neurodegeneration in adult rats: postnatal exposure enhances the susceptibility during adulthood. *Neurobiol Aging* 33:404–415
  96. Singh AK, Tiwari MN, Dixit A, Upadhyay G, Patel DK, Singh D, Prakash O, Singh MP (2011) Nigrostriatal proteomics of cypermethrin-induced dopaminergic neurodegeneration: microglial activation dependent and independent regulations. *Toxicol Sci* 122:526–538
  97. Tiwari MN, Singh AK, Ahmad I, Upadhyay G, Singh D, Patel DK, Singh C, Prakash O, Singh MP (2010) Effects of cypermethrin on monoamine transporters, xenobiotic metabolizing enzymes and lipid peroxidation in the rat nigrostriatal system. *Free Radic Res* 44:1416–1424
  98. Bachurin SO, Shevtsova EP, Kireeva EG, Oxenkrug GF, Sablin SO (2003) Mitochondria as a target for neurotoxins and neuroprotective agents. *Ann N Y Acad Sci* 993:334–344
  99. Miller JW, Selhub J, Joseph JA (1996) Oxidative damage caused by free radicals produced during catecholamine autoxidation: protective effects of O-methylation and melatonin. *Free Radic Biol Med* 21:241–249
  100. Sandyk R (1990) Pineal melatonin functions: possible relevance to Parkinson's disease. *Int J Neurosci* 50:37–53

101. Willis GL (2008) Intraocular microinjections repair experimental Parkinson's disease. *Brain Res* 1217:119–131
102. Reiter RJ, Carneiro RC, Oh CS (1997) Melatonin in relation to cellular antioxidative defense mechanisms. *Horm Metab Res* 29:363–372
103. Sandyk R (1990) The significance of eye blink rate in Parkinsonism: a hypothesis. *Int J Neurosci* 51:99–103
104. Dowling GA, Mastick J, Colling E, Carter JH, Singer CM, Aminoff MJ (2005) Melatonin for sleep disturbances in Parkinson's disease. *Sleep Med* 6:459–466
105. Tan DX, Reiter RJ, Manchester LC, Yan MT, El-Sawi M, Sainz RM, Mayo JC, Kohen R, Allegra M, Hardeland R (2002) Chemical and physical properties and potential mechanisms: melatonin as a broad spectrum antioxidant and free radical scavenger. *Curr Top Med Chem* 2:181–197
106. Khaldy H, Escames G, León J, Vives F, Luna JD, Acuña-Castroviejo D (2000) Comparative effects of melatonin, L-deprenyl, Trolox and ascorbate in the suppression of hydroxyl radical formation during dopamine autooxidation in vitro. *J Pineal Res* 29:100–107
107. Khaldy H, Escames G, Leon J, Bikjdaouene L, Acuña-Castroviejo D (2003) Synergistic effects of melatonin and deprenyl against MPTP-induced mitochondrial damage and DA depletion. *Neurobiol Aging* 24:491–500
108. Chakravarty S, Rizvi SI (2011) Circadian modulation of sodium–potassium ATPase and sodium–proton exchanger in human erythrocytes: in vitro effect of melatonin. *Cell Mol Biol (Noisy-le-grand)* 57:80–86
109. Chakravarty S, Rizvi SI (2011) Day and night GSH and MDA levels in healthy adults and effects of different doses of melatonin on these parameters. *Int J Cell Biol* 2011:404591
110. Weinreb O, Mandel S, Youdim MB (2003) Gene and protein expression profiles of anti- and pro-apoptotic actions of dopamine, R-apomorphine, green tea polyphenol (–)-epigallocatechin-3-gallate, and melatonin. *Ann N Y Acad Sci* 993:351–361
111. Lezoualc'h F, Sparapani M, Behl C (1998) N-acetyl-serotonin (normelatonin) and melatonin protect neurons against oxidative challenges and suppress the activity of the transcription factor NF-kappaB. *J Pineal Res* 24:168–178
112. Roy D, Belsham DD (2002) Melatonin receptor activation regulates GnRH gene expression and secretion in GT1-7 GnRH neurons. Signal transduction mechanisms. *J Biol Chem* 277:251–258
113. Kim D, Chung J (2002) Akt: versatile mediator of cell survival and beyond. *J Biochem Mol Biol* 35:106–115
114. Kong PJ, Byun JS, Lim SY, Lee JJ, Hong SJ, Kwon KJ, Kim SS (2008) Melatonin induces Akt phosphorylation through melatonin receptor- and PI3K-dependent pathways in primary astrocytes. *Korean J Physiol Pharmacol* 12:37–41
115. Lee SH, Chun W, Kong PJ, Han JA, Cho BP, Kwon OY, Lee HJ, Kim SS (2006) Sustained activation of Akt by melatonin contributes to the protection against kainic acid-induced neuronal death in hippocampus. *J Pineal Res* 40:79–85
116. Kilic U, Kilic E, Reiter RJ, Bassetti CL, Hermann DM (2005) Signal transduction pathways involved in melatonin-induced neuroprotection after focal cerebral ischemia in mice. *J Pineal Res* 38:67–71
117. Sharma R, McMillan CR, Tenn CC, Niles LP (2006) Physiological neuroprotection by melatonin in a 6-hydroxydopamine model of Parkinson's disease. *Brain Res* 1068:230–236
118. Niles LP, Armstrong KJ, Rincón Castro LM, Dao CV, Sharma R, McMillan CR, Doering LC, Kirkham DL (2004) Neural stem cells express melatonin receptors and neurotrophic factors: colocalization of the MT1 receptor with neuronal and glial markers. *BMC Neurosci* 5:41
119. Armstrong KJ, Niles LP (2002) Induction of GDNF mRNA expression by melatonin in rat C6 glioma cells. *Neuroreport* 13:473–475
120. Adi N, Mash DC, Ali Y, Singer C, Shehadeh L, Papapetropoulos S (2010) Melatonin MT1 and MT2 receptor expression in Parkinson's disease. *Med Sci Monit* 16:BR61–BR67
121. Buscemi N, Vandermeer B, Hooton N, Pandya R, Tjosvold L, Hartling L, Baker G, Klassen TP, Vohra S (2005) The efficacy and safety of exogenous melatonin for primary sleep disorders. A meta-analysis. *J Gen Intern Med* 20:1151–1158
122. Buscemi N, Vandermeer B, Hooton N, Pandya R, Tjosvold L, Hartling L, Vohra S, Klassen TP, Baker G (2006) Efficacy and safety of exogenous melatonin for secondary sleep disorders and sleep disorders accompanying sleep restriction: meta-analysis. *BMJ* 332:385–393
123. Bubenik GA, Konturek SJ (2011) Melatonin and aging: prospects for human treatment. *J Physiol Pharmacol* 62:13–19
124. Lauterbach EC, Victoroff J, Coburn KL, Shillcutt SD, Doonan SM, Mendez MF (2010) Psychopharmacological neuroprotection in neurodegenerative disease: assessing the preclinical data. *J Neuropsychiatry Clin Neurosci* 22:8–18
125. Schernhammer E, Chen H, Ritz B (2006) Circulating melatonin levels: possible link between Parkinson's disease and cancer risk? *Cancer Causes Control* 17:577–582
126. Zesiewicz TA, Evatt ML (2009) Potential influences of complementary therapy on motor and non-motor complications in Parkinson's disease. *CNS Drugs* 23:817–835
127. Sharma R, Mcmillan CR, Niles LP (2007) Neural stem cell transplantation and melatonin treatment in a 6-hydroxydopamine model of Parkinson's disease. *J Pineal Res* 43:245–254
128. Acuna Castroviejo D, López LC, Escames G, Lopez A, Garcia JA, Reiter RJ (2011) Melatonin–mitochondria interplay in health and disease. *Curr Top Med Chem* 11:221–240
129. Jou MJ, Peng TI, Hsu LF, Jou SB, Reiter RJ, Yang CM, Chiao CC, Lin YF, Chen CC (2010) Visualization of melatonin's multiple mitochondrial levels of protection against mitochondrial Ca(2+)-mediated permeability transition and beyond in rat brain astrocytes. *J Pineal Res* 48:20–38
130. Suzen S (2006) Recent developments of melatonin related antioxidant compounds. *Comb Chem High Throughput Screen* 9:409–419
131. Chiueh CC, Andoh T, Lai AR, Lai E, Krishna G (2000) Neuroprotective strategies in Parkinson's disease: protection against progressive nigral damage induced by free radicals. *Neurotox Res* 2:293–310